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2	HPV Oncogenic mRNA testing for Cervical Cancer Screening; Baseline and Longitudinal
3	Results from the CLEAR Study
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#### 77 ABSTRACT

78 This study determined the longitudinal clinical performance of a HR-HPV E6/E7 RNA assay 79 (Aptima HPV) compared to a HR-HPV DNA assay (Hybrid Capture 2) as an adjunctive method 80 for cervical cancer screening. Women  $\geq$  30 years with NILM cytology (n=10,860) positive by AHPV and/or HC2 assays, and randomly-selected women negative by both assays, were referred 81 to colposcopy at baseline. Women without baseline CIN2+ continued into 3-year follow-up. The 82 specificity of AHPV for <CIN2 was significantly greater at 96.3% compared to HC2 specificity 83 of 94.8% (p<0.001). Estimated sensitivities and risks for detection of CIN2+ were similar 84 85 between the two assays. After 3-years of follow-up, women negative by either HPV test had a 86 very low risk for CIN2+ (<0.3%) compared to CIN2+ risk in women with positive AHPV results 87 (6.3%) or positive HC2 results (5.1%). These results support the use of Aptima HPV as a safe 88 and effective adjunctive cervical cancer screening method.

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#### 91 INTRODUCTION

92 Cervical cancer is one of the most frequent cancers in women worldwide, accounting for 93 approximately 530,000 new cases and 275,000 deaths annually (1). Countries with well-94 organized screening programs using conventional Papanicolaou stain cytology have experienced 95 substantially reduced mortality from the disease in the past five decades (2-4). Despite this 96 advance, the relatively low sensitivity and reproducibility of both conventional Pap smear and liquid-based cytology (LBC) screening methods prompts investigation into identifying 97 adjunctive methods with Pap cytology for improving detection of cervical neoplasia (5-9). 98 Infection with 14 high-risk human papillomavirus (HR-HPX) genotypes (16, 18, 31, 33, 35, 39, 99 100 45, 51, 52, 56, 58, 59, 66, 68) is associated with almost all cases of cervical precancer, defined as 101 cervical intraepithelial neoplasia grade 2 (CIN2), grade 3 (CIN3), and cancer (10). Addition of 102 HR-HPV nucleic acid testing to a cervical cytology screening regimen offers higher sensitivity and negative predictive value (NPV) for detection of cervical precancer and cancer compared to 103 cytology alone, especially in older women (11-15). For this reason, HR-HPV nucleic acid testing 104 105 is recommended as an adjunctive test to cytology to assess the presence of HR-HPV types in 106 women 30 years of age or older (16). In this context, HR-HPV testing guides patient 107 management by identifying women at elevated risk for CIN2+, but importantly also reassures 108 women who are negative for HR-HPV of their extremely low cancer risk (17-19). 109 First generation HR-HPV molecular tests used for adjunctive cervical cancer screening function

by detecting viral genomic DNA in cellular samples from the uterine cervix. However, because the presence of HR-HPV in the female genital tract is common and often transient in nature (20-21), and most cervical HPV infections resolve without becoming cancerous (22-23), HR-HPV

113 DNA-based test methods yield only moderate specificity for detection of high-grade cervical

114 disease (12, 24). This leads to unnecessary follow-up and referral of patients to colposcopy,

115 increasing the physical and emotional burdens on patients and elevating health care costs.

116 An FDA-approved test for detection of HR-HPV E6/E7 mRNA (Aptima HPV Assay, AHPV)

has shown higher specificity with similar sensitivity for detection of CIN2+ as compared to HPV
DNA-based tests, in patients referred for colposcopy due to an abnormal Pap smear result as well

as in a screening setting (25-30). Expression of mRNA from viral E6 and E7 oncogenes is highly

120 associated with the development of cervical intraepithelial lesions (CIN) (31, 32), and extensive

121 investigation into the role of E6 and E7 oncoproteins in the HPV life cycle has revealed the

122 expression of the corresponding oncogenes is necessary and sufficient for cell immortalization,

123 neoplastic transformation, and the development of invasive cancer (33-35).

To confirm and extend the previous evidence on the clinical utility of HR-HPV oncogenic
mRNA testing in a U.S. population-based setting, the clinical performance of AHPV was
evaluated as an adjunctive method for cervical cancer screening in women aged 30 years or older
with NILM (negative for intraepithelial lesions or malignancy) cytology results from routine Pap
testing in a pivotal, prospective, multicenter U.S. clinical study including 3 years of follow-up
(the CLEAR [Clinical Evaluation of Aptima mRNA] study). We report herein the results from
this study.

#### 131 MATERIALS AND METHODS

#### 132 Study Design, Conduct and Participants

133 The CLEAR study consisted of 2 parts: the ASC-US (Atypical Squamous Cells of Unknown

134 Significance) Study (30) and the Adjunct Study described here (Fig. 1). Women 30 years of age

and older undergoing routine Pap testing who had a NILM cytology result were eligible to

136 participate in the Adjunct Study and were recruited from 19 US family planning and

137 obstetric/gynecologic clinics (private and academic), family practice medical groups, and clinical

138 research centers encompassing a wide geographic area representative of the US population.

139 Informed consent was obtained prior to enrollment of subjects. The study protocol was approved

by IRBs at the participating centers and the study was conducted in accordance with applicable

141 regulatory requirements and good clinical practices.

Women were excluded from the study if they were pregnant, were vaccinated against HPV, had 142 143 a history of cervical disease (cancer or precancerous) or an abnormal Pap test result in the 144 previous 12 months, or had a history of illness that could interfere with the study or create an 145 unacceptable risk to the subject. Demographic information and relevant medical information (cervical cancer history, prior HPV diagnosis, and any abnormal cytology history) were collected 146 from each subject. The study employed a baseline evaluation and a 3-year follow-up period with 147 annual cytology visits for longitudinal disease ascertainment. Subjects completed and exited the 148 study once they had a CIN2+ diagnosis. 149

150 Cytology (Referral Pap)

At the baseline evaluation and each annual visit thereafter, a cervical specimen was collected with a broom-like device (Papette; Wallach Surgical Devices, Orange, Conn) or an endocervical brush and spatula (Cytobrush Plus GT and Pap Perfect Plastic Spatula; Medscand, Trumbull, Conn) and placed into a ThinPrep Pap Test vial containing PreservCyt Solution ("referral Pap" specimen). Pap specimens were processed locally using the ThinPrep 2000 System (Hologic, Inc., Bedford, Mass) and evaluated for cytologic abnormalities. Cytology results were classified using the 2001 Bethesda System for reporting cervical cytology (36).

#### 158 HPV Testing

159 Baseline PreservCyt specimens (1 mL aliquot) were tested with the Aptima HPV Assay (AHPV;

- 160 Hologic, Inc., San Diego, California) on both the automated Tigris DTS System and Panther
- 161 System. Results from the two systems were similar; Panther System results are presented here.
- 162 AHPV is a target amplification assay that uses Transcription-Mediated Amplification (TMA) to
- 163 detect the E6/E7 oncogene mRNA of 14 HR-HPV genotypes (16, 18, 31, 33, 35, 39, 45, 51, 52,
- 164 56, 58, 59, 66, 68). Three clinical laboratories each tested approximately one-third of all samples
- 165 with AHPV. The majority of the PreservCyt specimens were also tested at one laboratory for
- 166 HR-HPV DNA using the Hybrid Capture 2 assay (HC2; Qiagen, Gaithersburg, Maryland), a
- 167 FDA-approved test that detects HR-HPV DNA of 13 of the 14 HR-HPV types detected by
- 168 AHPV and is known to cross react with the  $14^{th}$  type (37). Testing and results interpretation of
- 169 both HR-HPV tests were done according to manufacturer's instructions (38, 39). Technicians
- 170 performing HC2 and AHPV assays were masked to the other HPV test results and the subjects'
- 171 clinical status and colposcopic/histology results.
- 172 Disease Ascertainment

At baseline, women who tested positive in either the AHPV or the HC2 assay (HPV positive
women) and, to adjust for verification bias, approximately 6% of women who tested negative in
both HPV assays (HPV negative women) were randomly selected and referred to colposcopy.
Most colposcopy visits (>60%) were completed within 16 weeks from the baseline visit (median:
14 weeks; IQR= 8 weeks).

- 178 Colposcopists were masked to HPV results and collected cervical punch biops(ies) from each
- 179 visible lesion ("directed" biopsy) and an endocervical curettage (ECC) biopsy. The biopsy
- 180 specimens were processed according to the normal site procedures to produce H&E

181 (hemotoxylin and eosin) stained slides. After local pathologist review, slides were reviewed by 182 two central panel pathologists and classified using the three-tiered CIN terminology (40). Slides 183 with discordant central panel diagnoses were reviewed by a third central pathologist to reach a 184 consensus diagnosis (2 out of 3 agreement). If agreement was not achieved, the 3 central panel 185 pathologists reviewed the slides in conference to reach consensus. A subject's cervical disease 186 status represents the highest grade consensus histology result from colposcopy biopsy. Review 187 pathologists were masked to all other pathologists' diagnoses, the subjects' clinical status, enrollment status (ASC-US Study or Adjunct Study), and HPV test results. 188 189 During follow-up, women with ASC-US or more severe cytology results were referred to

colposcopy. Colposcopists collected the same types of biopsies which were processed and
submitted to the same central pathology review as done at baseline to obtain the consensus
histology result for that visit. Women with ASC-US or more severe cytology results who did not
have a colposcopy were considered to have indeterminate disease status. Subjects with NILM
cytology at a follow-up visit were not referred to colposcopy and were considered to have a
normal cervix.

The final disease status after 3-year follow-up was determined for each subject that completed 196 197 the study. To complete the study, a women must have either 1) a consensus result of CIN2 or 198 worse or 2) at least one cytology visit during the first or second year of follow-up and one 199 cytology visit during the third year of follow-up including colposcopy(ies) for those with ASC-200 US or more severe cytology. Final disease status for women meeting the second criteria was 201 based on their final consensus histology result or they were considered normal if they had NILM 202 cytology at the last visit. Women with CIN2 or worse did not have further follow-up in the 203 study.

Subjects with an ASC-US or more severe cytology during follow-up who did not have a
colposcopy or who attended the colposcopy visit but biopsies were not collected, were lost, or
the slides were inadequate to determine disease status were classified as indeterminate for
cervical disease status.

#### 208 Statistical Analysis

209 Test performance was evaluated with subjects having a consensus histology result of CIN2+

210 (CIN2, CIN3, carcinoma in situ or invasive cancer) classified as positive for cervical disease. A

211 diagnosis of CIN1 or normal disease status classified subjects as negative for cervical disease. In

addition, test performance was evaluated using a more definitive disease end point where a

consensus histology result of CIN3+ (CIN3, carcinoma in situ or invasive cancer) classified
subjects as positive for cervical disease and CIN2, CIN1, or normal classified subjects as

215 negative for cervical disease.

For the baseline risk analysis, a disproportionately smaller subset (3.4%) of HPV negative versus positive women had disease status determined from the baseline colposcopy visit, resulting in verification bias. To adjust for this bias, a multiple imputation method (41) was used to impute missing disease status based on the observed consensus histology results and AHPV and HC2 assay results from women who had a baseline colposcopy. Verification-bias adjusted risk estimates and 95% confidence intervals were generated using these imputed results.

222 The follow-up risk analysis included disease identified at baseline and during follow-up.

223 Because all women did not have colposcopy at baseline, individual by-year risk estimates may

reflect disease that was either present but not detected at baseline, or incident or progressive

disease. Cumulative risks with 95% CI were generated using the life-table method with subjects

not completing the study censored after the follow-up year last attended.

227 Sensitivity and specificity estimates with 95% CI were generated including women who

228 completed the study. McNemar's exact test of discordant matched pairs was performed to

compare the assays, including only subjects with results for both assays. All statistical tests and

230 CIs were 2-tailed and performed at the 5% significance level, using SAS<sup>®</sup> Version 9.1 or higher.

231 **RESULTS** 

#### 232 Subject Disposition and Demographic Information

A total of 13,495 women were included in this clinical study (Figure 1). Of the 12,869 women 30 233 years of age or older, 227 had an unsatisfactory or missing cytology result and 1001 had an 234 abnormal Pap result: ASC-US (5.7%), LSIL (1.5%), HSIL (0.2%), ASC-H (0.1%), or AGC, 235 236 AGC-favor neoplastic, or "other" (0.2% combined prevalence). The remaining 11,641 women  $\geq$ 30 years old with NILM cytology at baseline were enrolled in the Adjunct Study and tested 237 with the AHPV and HC2 tests. In total, 10,860 women were available for the baseline analysis, 238 including 864 women (525 HPV+ and 339 HPV-) with a baseline colposcopy (781 women 239 withdrew, see Figure 1 for reasons). Approximately 50% of the women referred to colposcopy 240 had an ECC biopsy only, and approximately 50% had ECC plus one or more directed biopsies, 241 242 resulting in the identification of 20 cases of CIN2+.

After the baseline evaluation, 10,509 women were eligible for follow-up (331 women withdrew for various reasons; see Fig. 1). During follow-up, 7,247 women returned for an annual cytology visit during Year 1, 6,517 returned during Year 2 and 6,339 returned during Year 3, with 6,201 women completing the study. Of the women who completed the study, 4452 returned during all 3 years; the remaining returned only once during the first 2 years and in Year 3, or had CIN2+ and exited the study prior to Year 3. In each follow-up year, 4-6% of the women had ASC-US or greater cytology.

- 250 Demographics are presented in Table 1. The median age was 43 years, with 61.4% age 40 years
- 251 or older; 44.0% were White Non-Hispanic, 16.6% were White-Hispanic, 12.5% were Black,
- 252 5.7% were Asian, and 21.1% were categorized as "Other" race or unknown.

#### 253 HPV and Disease Prevalence

- Cervical disease and HPV status are shown in Table 2 for the baseline evaluation (Table 2A) and cumulatively after 3 years of follow up (Table 2B). Of the 10,860 evaluable subjects with NILM
- cytology at baseline, 512 were positive for AHPV, yielding a prevalence of 4.7% for HR HPV
- 257 E6/E7 oncogenic mRNA, whereas prevalence of HR HPV DNA was 6.5% among 10,229
- women with HC2 results. A total of 845 HPV RNA-positive or DNA-positive women, and 556
- randomly-selected HPV-negative women, were referred to colposcopy at baseline (Figure 1).
- 260 At baseline, the percentage of colposcopy attendance was similar between HPV-positive (62%,
- n=526) and randomly-selected HPV-negative (61%, n=339) women with 29 CIN1, 9 CIN2, 8
- 262 CIN3, and 3 adenocarcinoma in situ (AIS) identified (Table 2A). Four of the CIN2 and two of
- the AIS cases were identified based on an ECC biopsy only.
- In total, 6,201 women completed the 3-year follow-up with a known disease status (Table 2B).
- 265 Of these, 6,098 (98.3%) women had normal (negative) disease status and 56 (0.9%) had low-
- grade lesions (CIN1). In addition to the 20 women with CIN2+ identified at baseline, 15 (0.2%)
- women had CIN2 and 12 (0.2%) women had CIN3 identified during follow-up, with two cases
- identified from an ECC biopsy only.
- 269 Of the 27 women with CIN2+ identified during follow-up, two had CIN1 at baseline with CIN3
- 270 identified during year 1. Ten women had no disease found at baseline with five CIN2+
- identified during year 1, one CIN2+ identified during year 2, and four CIN2+ identified during

272 year 3. The remaining 15 women with CIN2+ identified during follow-up did not have a baseline
273 colposcopy; among them, two CIN2+ were identified during year 1, six CIN2+ during year 2,

and seven CIN2+ during year 3.

#### 275 AHPV Assay Performance

276 Baseline risk and prevalence estimates adjusted for verification bias are provided in Table 3. The prevalence of CIN2+ was 0.9% in the overall population. CIN2+ occurred in 4.5% (95% CI: 277 2.7%, 7.4%) of women with positive AHPV results, and in 0.6% (95% CI: 0.2%, 1.9%) of 278 women with negative AHPV results, yielding a relative risk of 7.5 (95% CI: 2.1, 26.3). This 279 indicates that women with a positive AHPV result are at significantly greater risk of CIN2+ than 280 281 women with a negative AHPV result. The CIN2+ relative risk obtained for the HC2 test at baseline was similar (7.3; 95% CI: 1.6, 33.5). For CIN3+ diagnosis the overall prevalence was 282 283 0.4%. The AHPV relative risk was 24.9 (95% CI: 2.0, 307.0443.3), again with a similar relative

284 risk for HC2 (21.0; 95% CI: 1.0, 423,8).

Cumulative absolute and relative risks for AHPV and HC2 over the 3-year follow-up period for 285 HPV-positive and HPV-negative women are shown in Table 4. Women with an HPV-negative 286 287 result with either test had very low cervical disease risk after 3-years of follow-up (<0.3%). 288 Comparatively, 5-6% of women with an HPV-positive result had CIN2+ and 3-4% had CIN3+, 289 with overall cumulative absolute and relative risks slightly higher for the AHPV assay than for 290 HC2. Younger women aged 30-39 years who were HPV-positive had twice the prevalence of 291 disease but a similar increase in relative risk of cervical disease, compared to HPV-positive 292 women aged 40 years and older (Table 4). Risk of cervical disease in HPV-negative women did 293 not vary by age group.

- Figures 2 and 3 show the cumulative absolute risk of CIN2+ and CIN3+, respectively, by year
- according to AHPV or HC2 positivity status at baseline. Both assays show similar trend, with
- 296 consistent slightly higher risk for the AHPV assay each year.
- After 3-years of follow-up, the specificity of AHPV for <CIN2 was 96.3% (95% CI: 95.8%,
- 298 96.7%), significantly greater (p<0.001) compared to HC2 specificity of 94.8% (95% CI: 94.3%,
- 299 95.4%) (Table 5). AHPV specificity for <CIN3 (96.2% (95% CI: 95.5%, 96.5%)) was also
- 300 significantly greater (p<0.001) than HC2 specificity (94.7% (95% CI: 94.1%, 95.2%)).
- 301 Estimated sensitivities for detection of CIN2+ and CIN3+ were similar between the two assays
- 302 (p=0.219, p=1.0, respectively). For detection of CIN2+, AHPV sensitivity was 55.3% (95% CI:
- 303 41.2, 68.6), and HC2 sensitivity was 63.6% (95% CI: 48.9, 76.2). For CIN3+ detection, AHPV
- 304 sensitivity was 78.3% (95% CI: 58.1, 90.3), and HC2 sensitivity was 81.8% (95% CI: 61.5,
- 305 92.7). (Table 5).

#### 306 COMMENT

This study presents the results of a three-year longitudinal evaluation of AHPV as an adjunctive 307 method for screening women 30 years and older who have NILM Pap cytology results. 308 309 Consistent with previously published data (28, 29), these results demonstrate that HR-HPV 310 oncogenic E6/E7 mRNA testing has a sensitivity similar to a HR-HPV DNA-based test for detection of CIN2+ and CIN3+, and slightly, but significantly improved, specificity compared to 311 312 HR-HPV DNA testing for both endpoints. We found use of AHPV as an adjunctive method for 313 HPV-induced cervical disease screening provided disease detection capability similar to HC2 314 while reducing the false positive rate (from 5.2% to 3.7%) relative to the HPV DNA-based test. 315 Reduction of HPV detection in women without cervical disease minimizes the anxiety and 316 burden associated with spurious positive HPV molecular test results in women with NILM

317 cytology, decreases healthcare costs, and reduces unnecessary follow-up procedures, thereby

318 improving the safety of cervical cancer screening [unnecessary colposcopy is considered to be a

319 significant "harm" in the recent American Cancer Society guidelines (16)].

320 Importantly, we show that women with a NILM cytology result who also had a positive AHPV

321 result are approximately 24 times more likely to have CIN2+ disease after three years than

322 women with a negative AHPV result. This risk increased to approximately 68-fold for detection

323 of CIN3+ disease. Similar but slightly lower risk estimates were obtained with HC2,

demonstrating comparable accuracy of AHPV and HC2 for identifying subjects with CIN2+ and

325 CIN3+ in this respect.

After 3 years of follow-up, women in this study who were HPV-negative at baseline using any 326 327 test method had very low risk for CIN2+ (<0.3%), a result similar to previously published studies with HC2 (42, 43). These findings reinforce evidence from previous studies showing that HR-328 HPV nucleic acid testing should be performed as an adjunctive test to routine Pap for cervical 329 cancer screening of women aged 30 years or older to increase sensitivity of disease detection 330 (28). Correspondingly, compared to annual cytology-only screening, this study supports longer 331 332 screening intervals for women negative for both abnormal cytology and HPV E6/E7 mRNA, due 333 to the high NPV and low risk of disease afforded by this screening algorithm for three years 334 following a test-negative baseline visit. Extension of cervical cancer screening intervals 335 following negative HPV and cytology test results in women 30 years or older is a key 336 recommendation of current U.S. screening guidelines from both the American Cancer Society 337 and the U.S. Preventive Services Task Force (16).

Conversely, since the PPV of any HPV test in women with NILM cytology is low, additional
AHPV testing to detect persistent HR-HPV infection during follow-up care in women with an
initial AHPV positive result is likely a better option than direct referral to colposcopy.
Alternatively, genotyping with referral for HPV 16 or 18 positive women can optimize referral
and minimize loss to follow-up (44).

343 Several design features were employed in the CLEAR study to achieve accurate determination of the performance characteristics for both AHPV and HC2 assays. First, all biopsy samples were 344 subjected to adjudicated review by three independent expert pathologists. Second, molecular test 345 performance was compared to a consensus histology diagnosis, the gold standard for determining 346 347 cervical disease status. Third, AHPV performance was compared directly to HC2 performance, the most broadly used and characterized HPV DNA test. Fourth, performance characteristics of 348 349 both assays obtained from baseline results were adjusted for verification bias by conducting colposcopy and biopsy in 3.4% of HPV-negative women. This process is recommended in low 350 351 prevalence populations to avoid overestimating assay sensitivity and underestimating assay specificity (45-47). Finally, women were followed for 3 years with annual cytology testing and 352 referral to colposcopy for abnormal results. 353

A limitation of this study was that a portion of HPV-negative women with normal cytology were subjected to colposcopy and biopsy at the baseline visit but not at the subsequent follow up visits. Thus the relative risk estimates reported here for disease in HPV-positive vs –negative women evaluated during years 1, 2 and 3 of the follow up period may be overstated. This potential bias is present in previously reported longitudinal co-testing studies (17, 19, 48) and is unavoidable, since implementation of invasive procedures on thousands of women with normal cytology and negative HPV test results presents a burden to study subjects, and is not supported

by current US and European practice guidelines. However, as in previous longitudinal co-testing
studies, women enrolled in CLEAR who exited at the final (third) year of follow up had yielded
negative cytology and/or negative HC2 and AHPV results from 4 consecutive examinations.
Thus their risk of harboring an occult CIN lesion is likely to be exceedingly small (42, 43), such
that any potential error encountered here most likely constitutes a very small fraction of the
overall magnitude of the risks reported.

Another limitation of this study was that colposcopists were aware of the women's HPV test 367 368 status during the first half of the baseline portion of the study, because during that period, only 369 women who tested positive in AHPV or HC2 were referred to colposcopy. When the 370 colposcopists were unmasked, they may have been more diligent to find cervical disease with 371 prior knowledge of current HPV infection status. However, after randomly-selected HPVnegative women were referred to colposcopy, the colposcopists were masked to HPV status, and 372 throughout the entire study, colposcopists were masked as to which HPV assay caused the 373 374 referral. Thus any potential "colposcopy bias" would be identical for both molecular tests. 375 In summary, these results demonstrate the clinical performance of HR-HPV E6/E7 mRNA testing using AHPV is consistent with current U.S. cervical cancer screening guidelines for 376 377 women with a NILM cytology result who are  $\geq$ 30 years of age. There was a significantly greater risk of CIN2+ in AHPV-positive versus AHPV-negative subjects, as well as a statistically and 378 379 clinically significant improvement in specificity for detection of CIN2+ by AHPV compared to 380 HPV DNA testing with the HC2 assay. Thus, these data confirm the clinical utility of Aptima HPV testing in an adjunct cervical cancer screening setting. 381

#### 382 **REFERENCES**

383	1.	Jemal A, Bray F, Center M, et al. Global cancer statistics. CA Cancer J Clin 2011;61:69-
384		90.

Sasieni PD, Cuzick J, Lynch-Farmery E. Estimating the efficacy of screening by auditing
 smear histories of women with and without cervical cancer. Br J Cancer 1996; 73: 1001 5.

- Gustafsson L, Ponten J, Zack M, et al. International incidence rates of invasive cervical
   cancer after introduction of cytological screening. *Cancer Causes Control*. 1997;8:755 763
- 4. U.S. Cancer Statistics Working Group. United States Cancer Statistics: 2004 Incidence
   and Mortality. Atlanta: U.S. Department of Health and Human Services, Centers for
   Disease Control and Prevention and National Cancer Institute; 2007. Accessed at
   www.cdc.gov/cancer/npcrpdfs/US Cancer Statistics 2004 Incidence and Mortaliy.pdf
- 5. Franco E, Syrjanen K, de-Wolf C, et al. New developments in cervical cancer screening
  and prevention. Geneva, Switzerland, June 17-19, 1996. Workshop. Cancer Epidemiol
  Biomarkers Prev 1996;5:853-6.
- Miller AB, Nazeer S, Fonn S, et al. Report on consensus conference on cervical cancer
   screening and management. Int J Cancer 2000;86:440-7.

400 7. Stoler MH, Schiffman M. Atypical Squamous Cells of Undetermined Significance-Low401 grade Squamous Intraepithelial Lesion Triage Study (ALTS) Group. Interobserver
402 reproducibility of cervical cytologic and histologic interpretations: realistic estimates
403 from the ASCUS-LSIL Triage Study. JAMA 2001;285:1500-5.

# 404 8. Arbyn M, Bergeron C, Klinkhamer P, et al. Liquid compared with conventional cervical 405 cytology: a systematic review and meta-analysis. Obstet Gynecol 2008;111:167-77.

9. Syrjänen KJ, Shabalova IP, Ivanchenko O, et al. Reproducibility of classification and
correction for verification bias as determinants of performance of Papanicolaou smear
cytology in the screening setting: Experience from the New Independent States of the
former Soviet Union cohort study. Acta Cytologica 2009;53:548-57.

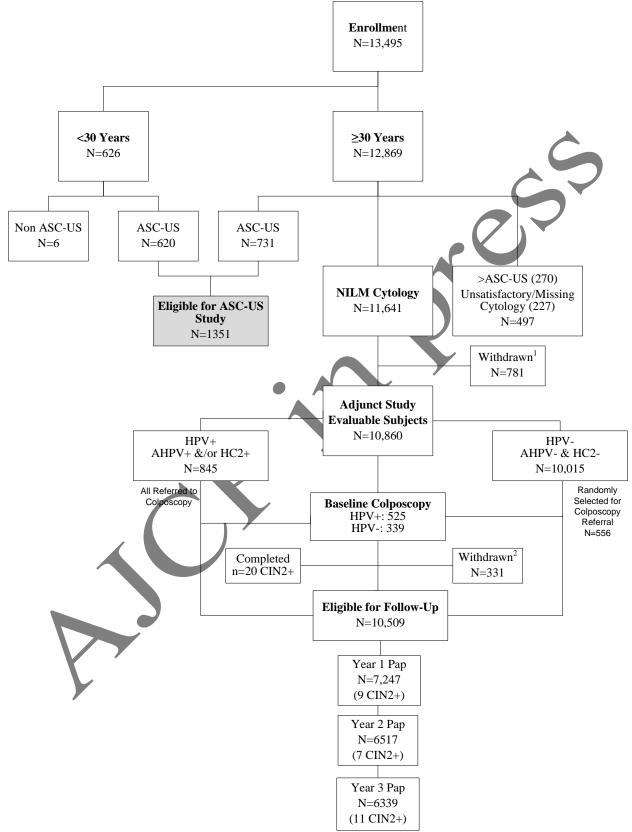
410	10. Walboomers JM, Jacobs MV, Manos MM, et al. Human papillomavirus is a necessary
411	cause of invasive cervical cancer worldwide. J Pathol. 1999 Sep;189(1):12-9.
412	11. Kulasingam SL, Hughes JP, Kiviat NB, et al. Evaluation of human papillomavirus testing
413	in primary screening for cervical abnormalities: comparison of sensitivity, specificity,
414	and frequency of referral. JAMA 2002;288:1749-57.
415	12. Cuzick, JC, Clavel KU, Petry CJ, et al. Overview of the European and North American
416	studies on HPV testing in primary cervical cancer screening. Int J Cancer 2006;
417	119:1095-101.
418	13. Mayrand MH, Duarte-Franco E, Rodrigues I, et al. Canadian Cervical Cancer Screening
419	Trial Study Group. Human papillomavirus DNA versus Papanicolaou screening tests for
420	cervical cancer. N Engl J Med 2007;357:1579-88.
421	14. Ronco G, Giorgi-Rossi P, Carozzi F, et al. New Technologies for Cervical Cancer
422	Screening Working Group. Results at recruitment from a randomized controlled trial
423	comparing human papillomavirus testing alone with conventional cytology as the primary
424	cervical cancer screening test. J Natl Cancer Inst 2008;100:492-501.
425	15. Baseman JG, Kulasingam SL, Harris TG, et al. Evaluation of primary cervical cancer
426	screening with an oncogenic human papillomavirus DNA test and cervical cytologic
427	findings among women who attended family planning clinics in the United States. Am J
428	Obstet Gynecol 2008;199:26-8.
429	16. Saslow D, Solomon D, Lawson H, et al. American Cancer Society, American Society for
430	Colposcopy and Cervical Pathology, and American Society for Clinical Pathology
431	Screening Guidelines for the Prevention and Early Detection of Cervical Cancer. Am J
432	Clin Pathol 2012;137:516-542.
433	17. Khan M , Castle PE , Lorincz AT, et al. The Elevated 10-Year Risk of Cervical Precancer
434	and Cancer in Women With Human Papillomavirus (HPV) Type 16 or 18 and the
435	Possible Utility of Type-Specific HPV Testing in Clinical Practice. J. Nat Cancer Inst.
436	2005; 97: 1072-1079.
437	18. Ronco G, Giorgi-Rossi P, Carozzi F, et al. New Technologies for Cervical Cancer
438	screening (NTCC) Working Group. Efficacy of human papillomavirus testing for the

- 439 detection of invasive cervical cancers and cervical intraepithelial neoplasia: a randomised
  440 controlled trial. Lancet Oncol. 2010 Mar;11(3):249-57
- 441 19. Rijkaart DC, Berkhof J, Rozendaal L,et al. Human papillomavirus testing for the
   442 detection of high-grade cervical intraepithelial neoplasia and cancer: final results of the
- 443 POBASCAM randomised controlled trial. Lancet Oncol. 2012 Jan;13(1):78-88.
- 20. Moscicki A-B, Ma Y, Jonte J, et al. The Role of Sexual Behavior and Human
  Papillomavirus Persistence in Predicting Repeated Infections with New Human
  Papillomavirus Types. 2010a; Cancer Epidemiol Biomarkers Prev19:2055-2065.
- 447 21. Goodman M, Shvetsov Y, McDuffie K, et al. Prevalence, Acquisition, and Clearance of
  448 Cervical Human Papillomavirus Infection among Women with Normal Cytology: Hawaii
  449 Human Papillomavirus Cohort Study. Cancer Res 2008; 68, 8813-8824.
- 450 22. Münger K, Baldwin A, Edwards KM, et al. Mechanisms of human papillomavirus451 induced oncogenesis. J Virol 2004;78:11451-60.
- 452 23. Moscicki A-B, Ma Y, Wibbelsman C, et al. Rate of and Risks for Regression of CIN-2 in
  453 adolescents and young women. Obstet Gynecol. 2010b December; 116(6): 1373–1380.
- 454 24. Koliopoulos G, Arbyn M, Martin-Hirsch P, et al. Diagnostic accuracy of human
  455 papillomavirus testing in primary cervical screening: A systematic review and meta456 analysis of non-randomized studies. Gynecol Oncol 2007;104:232-46.
- 457 25. Szarewski A, Ambroisine L, Cadman L, et al. Comparison of predictors for high-grade
  458 cervical intraepithelial neoplasia in women with abnormal smears. Cancer Epidemiol
  459 Biomarkers Prey 2008;17:3033-42.
- 26. Clad A, Reuschenbach M, Weinschenk J, et al. Performance of the Aptima high-risk
  human papillomavirus mRNA assay in a referral population in comparison with Hybrid
  Capture 2 and cytology. J Clin Microbiol. 2011 Mar;49(3):1071-6.
- 27. Ratnam S, Coutlee F, Fontaine D, et al. Aptima HPV E6/E7 mRNA test is as sensitive as
  Hybrid Capture 2 Assay but more specific at detecting cervical precancer and cancer. J
  Clin Microbiol. 2011 Feb;49(2):557-64.
- 466 28. Monsonego J, Hudgens MG, Zerat L, et al. Evaluation of oncogenic human
  467 papillomavirus RNA and DNA tests with liquid-based cytology in primary cervical

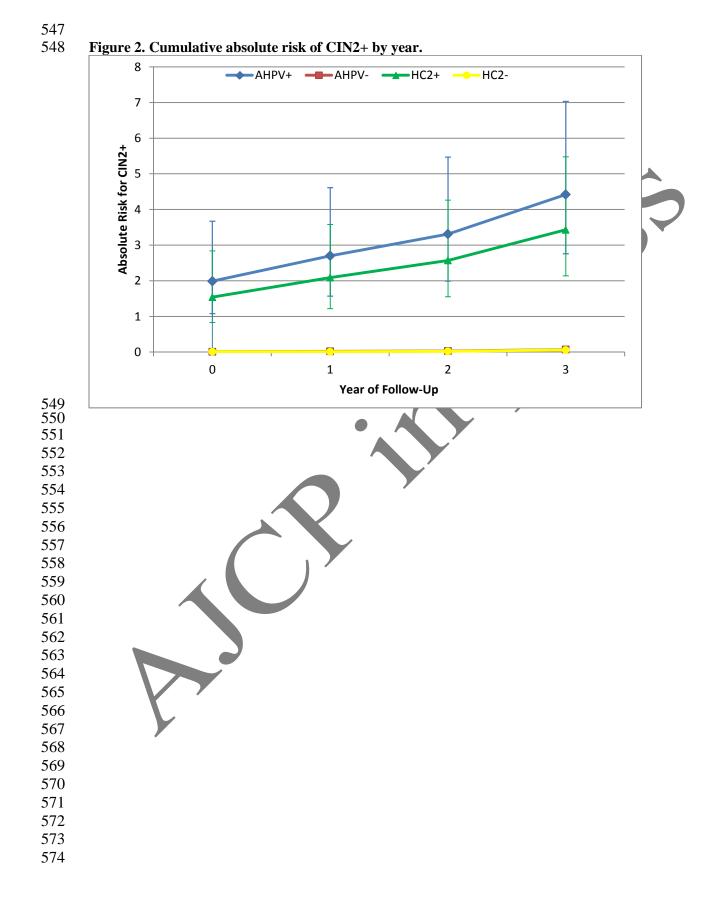
468	cancer screening: The FASE study. Int J Cancer. 2010; 129: 691-701.
469	29. Wu R, Belinson SE, Du H, et al. Human papillomavirus messenger RNA assay for
470	cervical cancer screening: the Shenzhen Cervical Cancer Screening Trial I. Int J Gynecol
471	Cancer. 2010 Nov;20(8):1411-4.
472	30. Stoler MH, Wright TC Jr, Cuzick J, et al. APTIMA HPV Assay Performance in Women
473	with Atypical Squamous Cells of Undetermined Significance Cytology Results. AJOG;
474	2013; 208 (2): 144-148.
475	31. zur Hausen H. Molecular pathogenesis of cancer of the cervix and its causation by
476	specific human papillomavirus types. Curr Top Microbiol Immunol 1994;186:131-56.
477	32. Sotlar K, Stubner A, Diemer D, et al. Detection of high-risk human papillomavirus E6
478	and E7 oncogene transcripts in cervical scrapes by nested RT-polymerase chain reaction.
479	J Med Virol 2004;74:107-16.
480	33. Klingelhutz, A. J., S. A. Foster, and J. K. McDougall. 1996. Telomerase activation by the
481	E6 gene product of human papillomavirus type 16. Nature 380:79–82.
482	34. Schreiber K, Cannon R, Karrison T, et al. Strong synergy between mutant ras and HPV16
483	E6/E7 in the development of primary tumors. Oncogene 2004; 23:3972–3979.
484	35. Liu X, Clements A, Zhao K, et al. Structure of the Human Papillomavirus E7
485	Oncoprotein and its mechanism for inactivation of the retinoblastoma tumor suppressor. J
486	Biol Chem, 2006; 281, 578-586.
487	36. Solomon D, Davey D, Kurman R, et al. The Bethesda System 2001: terminology for
488	reporting the results of cervical cytology. JAMA. 2002; 287:2114–2119.
489	37. Castle P, Solomon D, Wheeler CM, et al. Human Papillomavirus Genotype Specificity of
490	Hybrid Capture 2. J Clin Micro, Aug. 2008, p. 2595–2604.
491	38. Hologic, Inc APTIMA® HPV Assay [package insert]. San Diego, Calif; 2008.
492	39. Qiagen Hybrid Capture® 2 High-Risk HPV DNA Test® [package insert]. Gaithersburg,
493	Md; 2007.

494	40. Wright TC, Ronnett BM, Kurman RJ, et al. Precancerous lesions of the cervix. 2011. In
495	Blaustein's Pathology of the Female Genital Tract Sixth Edition. Editors Kurman RJ,
496	Ellenson LH, Ronnett BM. Springer New York, NY.
497	41. Little RJA and Rubin DB. Statistical analysis with missing data. 2nd ed. New York, NY:
498	John Wiley & Sons, Inc.; 2002:85-9.
499	42. Sherman ME, Lorincz AT, Scott DR. Baseline Cytology, Human Papillomavirus Testing,
500	and Risk for Cervical Neoplasia: A 10-Year Cohort Analysis. J Natl Cancer Inst. 2003
501	Jan 1;95(1):46-52.
502	43. Dillner J, Rebolj M, Birembaut P, et al. Joint European Cohort Study. Long term
503	predictive values of cytology and human papillomavirus testing in cervical cancer
504	screening: joint European cohort study. BMJ. 2008 Oct 13;337.
505	44. Castle PE, Cuzick J, Stoler M, et al. Detection of HPV16, 18, and 45 in Women with
506	ASC-US Cytology and the Risk of Cervical Precancer: Results from the CLEAR HPV
507	Study. Am J Clin Pathol, in press.
508	45. Zhou, X. Correcting for verification bias in studies of a diagnostic test's accuracy. Statist
509	Meth Med Res 1998;7:337-53.
510	46. Zhou X. Comparing accuracies of two screening tests in a two-phase study for dementia.
511	Appl Stat 1998; 47:135-47.
512	47. Begg CB, Greenes RA. Assessment of diagnostic tests when disease is subject to
513	selection bias. Biometrics 1983;39:207-16.
514	48. Katki H, Kinney W, Fetterman B, et al. Cervical cancer risk for women undergoing
515	concurrent testing for human papillomavirus and cervical cytology: a population-based
516	study in routine clinical practice. The Lancet Oncology. 2011; 12: 663-672.
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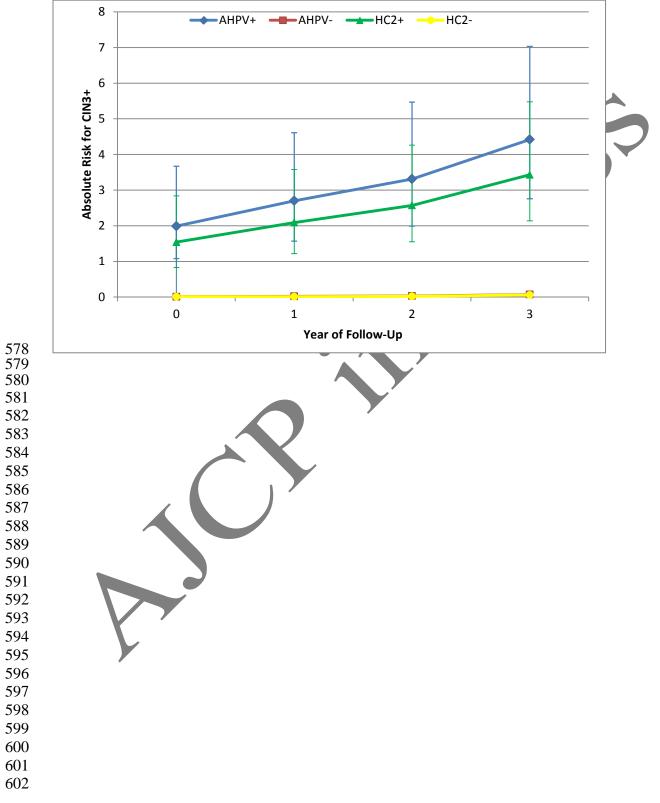




526	<sup>1</sup> Reasons for withdrawal: did not meet eligibility criteria (70); Pap volume insufficient for
527	AHPV testing (117); specimen expired or unsuitable for testing (190); specimen lost (58);
528	noncompliant site (320); other reasons (26).
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530	<sup>2</sup> Reasons for withdrawal: Collection site did not participate in follow-up (243); subject
531	terminated participation (37); subject had hysterectomy (22); subject not eligible (17); subject
532	treated prior to CIN2+ diagnosis (8); other reasons (4)
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## 577 Figure 3. Cumulative absolute risk of CIN3+ by year.



## **TABLES**

## **Table 1. Demographics of evaluable subjects**

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(A)Disease		AHPV+ (n=512)			AHPV- (n=10,348)		
Status at Baseline <sup>1</sup>	Subjects at Baseline N=10,860	HC2+ (n=383)	HC2- (n=97)	HC2 missing <sup>2</sup> (n=32)	HC2+ (n=282)	HC2- (n=9467)	HC2 missing (n=599)
Verified							
Normal	769	211	19	12	170	353	4
CIN1	29	12	0	1	7	9	0
CIN2	9	4	0	0	2	2	1
CIN3	8	7	0	0	1	0	0
AIS	3	2	1	0	0	0	0
CIN2+	20	13	1	0	3	2	1
CIN3+	11	9	1	0	1	0	0
Unverified	10,042	147	77	19	102	9103	594
(B)Disease Status After	All	AHPV+ (n=511)			AHPV- (n=10332)		
3-Year Follow-Up <sup>3</sup>	Subjects N=10,843 <sup>4</sup>	HC2+ (n=382)	HC2- (n=97)	HC2 missing <sup>2</sup> (n=32)	HC2+ (n=281)	HC2- (n=9452)	HC2 missing (n=599)
Normal	6098	161	48	10	123	5440	316
CIN1	56	10	0	0	6	36	4
CIN2	24	7	0	1	3	12	1
CIN3	20	14	0	1	2	3	0
AIS	3	2	1	0	0	0	0
CIN2+	47	23	1	2	5	15	1
CIN3+	23	16	1	1	2	3	0
Missing	4378	167	44	17	130	3756	264
Indeterminate	264	21	4	3	17	205	14

Table 2. Disease status at baseline (A) and after three years of follow-up (B) and corresponding AHPV and HC2 test results at baseline

histology result. Women without a consensus histology result have an unverified disease status. <sup>2</sup> 631 women with APTIMA HPV Assay results did not have HC2 test results primarily due to insufficient volume of the cytology specimen. <sup>3</sup> Disease status after 3-year follow-up is based on completing 3-year follow-up with cytology performed at least once and colposcopy attendance for  $\geq$ ASC-US results during the first 2 years and during the third year.

<sup>1</sup> Verified disease status was determined for women who attended colposcopy at baseline and had a consensus

<sup>4</sup> 17 women were determined ineligible after completion of baseline, results are excluded from follow-up analyses.

## 642 Table 3. Absolute and relative risk of CIN2+ and CIN3+ disease at baseline (verification-

**bias adjusted**)

Disease	Assay					
			HPV Assay	HC2 Test		
Status	Result	Absolute Risk (95% CI)	Relative Risk (95% CI)	Absolute Risk (95% CI)	Relative Ris (95% CI)	
≥CIN2	Positive	4.5 (2.7, 7.4)	7.5	3.7 (2.3, 6.1)	7.3	
	Negative	0.6 (0.2, 1.9)	(2.1, 26.3)	0.5 (0.1, 2.1)	(1.6, 33.5)	
	Prevalence (%)		9%		).9%	
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≥CIN3	Positive	3.0 (1.6, 5.5)	24.9	2.3 (1.3, 4.1)	21.0	
	Negative	0.1 (0.0, 1.7)	(2.0, 307.0)	0.1 (0.0, 2.4)	(1.0, 423.8)	
	Prevalence (%)	0.	4%	(	).4%	
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## Table 4. Cumulative absolute and relative risk of CIN2+ and CIN3+ disease by age group after 3-year follow-up (life-table analysis)

	APTIMA HPV Assay				HC2 Test		
	Disease Status	Age Group	Assay Result	Absolute Risk (95% CI)	Relative Risk (95% CI)	Absolute Risk (95% CI)	Relative Risk (95% CI)
	≥CIN2	Overall	Positive	6.32 (4.29, 9.27)	23.94	5.12 (3.53, 7.41)	22.39
			Negative	0.26 (0.17, 0.41)	(13.59, 42.18)	0.23 (0.14, 0.38)	(12.19, 41.12)
			Prevalence (%)	0.55	5%	0.55	%
		30 to 39	Positive	7.76 (4.81, 12.40)	31.11	6.46 (3.99, 10.39)	27.36
		Years	Negative Prevalence (%)	0.25 (0.12, 0.53) 0.76	(13.04, 74.21) 5%	0.24 (0.10, 0.54)	(10.88 - 68.80) %
		≥40	Positive	4.51 (2.34, 8.63)	16.57	3.77 (2.10, 6.71)	16.85
		Years	Negative	0.27 (0.16, 0.46)	(7.26, 37.82)	0.22(0.12, 0.42)	(7.21, 39.35)
			Prevalence (%)	0.42		0.40	
	≥CIN3	Overall	Positive	4.42 (2.76, 7.03)	67.87	3.43 (2.14, 5.48)	59.14
	<u>-</u> env5	Overall	Negative	4.42 (2.76, 7.03) 0.07 (0.03, 0.16)	(25.32, 181.88)	0.06 (0.02, 0.16)	(20.09 -174.12)
			Prevalence (%)	0.07 (0.03, 0.18) 0.27		0.08 (0.02, 0.18)	
		30 to 39	Positive	5.74 (3.22, 10.11)	102,84	4.78 (2.67, 8.48)	171.50
		Years	Negative	0.06 (0.01, 0.22)	(23.17, 456.51)	0.03 (0.00, 0.20)	(22.39 - 1313.63)
			Prevalence (%)	0.44	1%	0.45	
		≥40	Positive	2.81 (1.27, 6.16)	41.80	2.05 (0.93, 4.52)	28.46
		Years	Negative Prevalence (%)	0.07 (0.02, 0.21) 0.16	(10.53, 166.00)	0.07 (0.02, 0.22) 0.17	(7.15 - 113.20)
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